

This Page Is Inserted by IFW Operations
and is not a part of the Official Record

BEST AVAILABLE IMAGES

Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images may include (but are not limited to):

- BLACK BORDERS
- TEXT CUT OFF AT TOP, BOTTOM OR SIDES
- FADED TEXT
- ILLEGIBLE TEXT
- SKEWED/SLANTED IMAGES
- COLORED PHOTOS
- BLACK OR VERY BLACK AND WHITE DARK PHOTOS
- GRAY SCALE DOCUMENTS

IMAGES ARE BEST AVAILABLE COPY.

**As rescanning documents *will not* correct images,
please do not report the images to the
Image Problems Mailbox.**

FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AM	Armenia	GB	United Kingdom	MW	Malawi
AT	Austria	GE	Georgia	MX	Mexico
AU	Australia	GN	Guinea	NE	Niger
BB	Barbados	GR	Greece	NL	Netherlands
BE	Belgium	HU	Hungary	NO	Norway
BF	Burkina Faso	IE	Ireland	NZ	New Zealand
BG	Bulgaria	IT	Italy	PL	Poland
BJ	Benin	JP	Japan	PT	Portugal
BR	Brazil	KE	Kenya	RO	Romania
BY	Belarus	KG	Kyrgyzstan	RU	Russian Federation
CA	Canada	KP	Democratic People's Republic of Korea	SD	Sudan
CF	Central African Republic	KR	Republic of Korea	SE	Sweden
CG	Congo	KZ	Kazakhstan	SG	Singapore
CH	Switzerland	LI	Liechtenstein	SI	Slovenia
CI	Côte d'Ivoire	LK	Sri Lanka	SK	Slovakia
CM	Cameroon	LR	Liberia	SN	Senegal
CN	China	LT	Lithuania	SZ	Swaziland
CS	Czechoslovakia	LU	Luxembourg	TD	Chad
CZ	Czech Republic	LV	Latvia	TG	Togo
DE	Germany	MC	Monaco	TJ	Tajikistan
DK	Denmark	MD	Republic of Moldova	TT	Trinidad and Tobago
EE	Estonia	MG	Madagascar	UA	Ukraine
ES	Spain	ML	Mali	UG	Uganda
FI	Finland	MN	Mongolia	US	United States of America
FR	France	MR	Mauritania	UZ	Uzbekistan
GA	Gabon			VN	Viet Nam

DETECTION OF A LEPTIN RECEPTOR
VARIANT AND METHODS FOR REGULATING OBESITY

1. INTRODUCTION

5 The present invention relates to a variant form of the
receptor for the *obese* gene product. In particular, the
invention relates to methods of detecting this receptor
variant in cells and tissues of obese individuals. In
10 addition, it relates to methods of inhibiting or down-
regulating expression of this variant in cells to augment
their responsiveness to weight regulation by leptin as well
as methods of using compounds to directly activate signal
transduction pathways associated with this ligand-receptor
15 system.

2. BACKGROUND OF THE INVENTION

Obesity is not only a nutritional disorder in Western
societies, it is also a serious health concern because of its
20 association with adult-onset diabetes, hypertension, and
heart disease (Grundy, 1990, *Disease-a-Month* 36:645-696).
While there was evidence to suggest that body weight was
physiologically regulated, the molecular mechanism has
remained elusive. However, animal studies have produced
25 several mouse strains that contain single-gene mutations,
resulting in an obese phenotype. One such recessive mutation
is manifested in the *ob/ob* mice, and it is referred to as the
obese (ob) mutation.

Zhang et al. (1994, *Nature* 372:425-432) describe the
30 cloning and sequencing of the mouse *ob* gene and its human
homolog. When an isolated gene fragment was used as a probe,
it was shown to hybridize with RNA only in white adipose
tissue by northern blot analysis, but no expression was
detected in any other tissue. In addition, the coding
35 sequence of the *ob* gene hybridized to all vertebrate genomic
DNAs tested, indicating a high level of conservation of this
molecule among vertebrates. The deduced amino acid sequences

are 84% identical between human and mouse, and both molecules contain features of secreted proteins.

In an effort to understand the physiologic function of the *ob* gene, several independent research groups produced 5 recombinant *ob* gene product in bacteria for *in vivo* testing (Pelleymounter et al., 1995, *Science* 269:540-543; Halaas et al., 1995, *Science* 269:543-546; Campfield et al., 1995, *Science* 269:546-549). When the Ob protein (also known as leptin) was injected into grossly obese mice, which possessed 10 two mutant copies of the *ob* gene, the mice exhibited a reduced appetite and began to lose weight. In addition, these studies described a dual action of leptin in both reducing the animals' food intake and in increasing their energy expenditure. Similarly, when normal mice received 15 leptin, they also ate less than the untreated controls. More importantly, Campfield et al. (1995, *Science* 269:546-549) injected leptin directly into lateral ventricle, and observed a reduction in the animals' food intake, suggesting that leptin acts on central neuronal networks to regulate feeding 20 behavior and energy balance. Thus, this result provides evidence that the leptin receptor (also known as OB-R) is expressed by cells in the brain.

Recently, a leptin fusion protein was generated and used to screen for OB-R in a cDNA expression library prepared from 25 mouse choroid plexus, a tissue that lines brain cavities termed ventricles (Tartalia, 1995, *Cell* 83:1263-1271). This approach led to the cloning of one form of the OB-R coding sequence, which reveals a single membrane-spanning receptor, sharing structural similarities with several Class I cytokine 30 receptors, such as the gp130 signal-transducing component of the interleukin-6 receptor (Taga et al., 1989, *Cell* 58:573-581), the granulocyte-colony stimulating factor receptor (Fukunaga et al., 1990, *Cell* 61:341-350), and the leukemia inhibitory factor receptor (Gearing et al., 1991, *EMBO J.* 35 10:2839-2848). Northern blot analysis and reverse transcription-polymerase chain reaction (RT-PCR) demonstrate

that OB-R mRNA is expressed in several tissues, including lung, kidney, total brain, choroid plexus and hypothalamus.

The reported mouse OB-R protein contains a relatively short intracellular cytoplasmic domain as compared with other Class I cytokine receptors. Subsequently, when cDNA encoding its human homolog was isolated from a human infant brain library, the predicted human protein sequence contains a much longer intracellular domain. In view of this finding, it was speculated that different forms of the receptor might exist (Barinaga, 1996, *Science* 271:29). However, prior to the present invention, there was no report on the identification of any variant forms of the OB-R in humans or how such molecules, if they exist, would relate to obesity.

Additionally, several studies have shown that *ob* gene expression is actually increased in obese humans (Considine et al., 1995, *J. Clin. Invest.* 95:2986-2988; Lonquist et al., 1995, *Nature Med.* 1:950; Hamilton et al., 1995, *Nature Med.* 1:953). Moreover, the mutations in the mouse *Ob* gene were not detected in human mRNA. Therefore, taken collectively, these studies imply that decreased leptin levels are not the primary cause of obesity, and argue for the presence of a less responsive receptor in obese individuals. There remains a need to isolate such an OB-R variant for the design of therapeutics to augment weight regulation by leptin.

3. SUMMARY OF THE INVENTION

The present invention relates to a variant form of the human OB-R. In particular, it relates to the detection of this receptor variant in cells of obese individuals, and methods for treating obesity by targeting this variant.

The invention is based, in part, upon the Applicants' discovery of human cDNA clones encoding a variant form of the OB-R. This receptor differs structurally from a reported OB-R with only three amino acid substitutions in the extracellular domain, but extensive diversity is observed in their intracellular cytoplasmic domains at the 3' end. The

cytoplasmic domain of the variant of the invention is both shorter and distinct in nucleotide sequence from the corresponding domain of the published form of OB-R. Therefore, a wide variety of uses are encompassed by the present invention, including but not limited to, the detection of the receptor variant in cells of obese individuals, methods to inhibit and/or down-regulate the expression of this receptor variant, gene therapy to replace the receptor variant in homozygous individuals, and direct activation of downstream signal transduction pathways in cells expressing the receptor variant for weight regulation.

4. BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1A-1E. Nucleotide sequence (SEQ ID NO:1) and deduced amino acid sequence (SEQ ID NO: 2, 3 and 4) of the human OB-R variant. The amino acid sequence diverges from the human OB-R reported by Tartaglia et al. (1995, Cell 83:1263-1271) at nucleotide residue #349, #422, #764 and from residue #2770 and beyond.

5. DETAILED DESCRIPTION OF THE INVENTION

5.1. THE OB-R VARIANT

The present invention relates to nucleic acid and amino acid sequences of an OB-R variant in the Class I cytokine receptor family. In a specific embodiment by way of example in Section 6, *infra*, this variant was cloned and characterized. Amino acid sequence comparison of this OB-R variant with a published human OB-R sequence (Tartaglia et al., 1995, Cell 83:1263-1271) reveals three amino acid differences in their extracellular domain and extensive diversity in their intracellular cytoplasmic domains. More specifically, Figure 1A-1E shows that in the variant, nucleotide residues #349-351 encode alanine, nucleotide residues #421-423 encode arginine and nucleotide residues #763-765 encode arginine. Additionally, the variant diverges both in length and sequence composition from the published human OB-R sequence from nucleotide residue #2770 and beyond.

In order to clone additional variant forms of the molecule, labeled DNA probes made from nucleic acid fragments corresponding to any portion of the cDNA disclosed herein may be used to screen a cDNA library prepared from human fetal liver, human lung, human kidney, human choroid plexus and human hypothalamus. More specifically, oligonucleotides corresponding to either the 5' or 3' terminus of the cDNA sequence may be used to obtain longer nucleotide sequences. Briefly, the library may be plated out to yield a maximum of 30,000 pfu for each 150 mm plate. Approximately 40 plates may be screened. The plates are incubated at 37°C until the plaques reach a diameter of 0.25 mm or are just beginning to make contact with one another (3-8 hours). Nylon filters are placed onto the soft top agarose and after 60 seconds, the filters are peeled off and floated on a DNA denaturing solution consisting of 0.4N sodium hydroxide. The filters are then immersed in neutralizing solution consisting of 1M Tris HCL, pH 7.5, before being allowed to air dry. The filters are prehybridized in casein hybridization buffer containing 10% dextran sulfate, 0.5M NaCl, 50mM Tris HCL, pH 7.5, 0.1% sodium pyrophosphate, 1% casein, 1% SDS, and denatured salmon sperm DNA at 0.5 mg/ml for 6 hours at 60°C. The radiolabelled probe is then denatured by heating to 95°C for 2 minutes and then added to the prehybridization solution containing the filters. The filters are hybridized at 60°C for 16 hours. The filters are then washed in 1X wash mix (10X wash mix contains 3M NaCl, 0.6M Tris base, and 0.02M EDTA) twice for 5 minutes each at room temperature, then in 1X wash mix containing 1% SDS at 60°C for 30 minutes, and finally in 0.3X wash mix containing 0.1% SDS at 60°C for 30 minutes. The filters are then air dried and exposed to x-ray film for autoradiography. After developing, the film is aligned with the filters to select a positive plaque. If a single, isolated positive plaque cannot be obtained, the agar plug containing the plaques will be removed and placed in lambda dilution buffer containing 0.1M NaCl, 0.01M magnesium sulfate, 0.035M Tris HCL, pH 7.5, 0.01% gelatin. The phage

may then be replated and rescreened to obtain single, well isolated positive plaques. Positive plaques may be isolated and the cDNA clones sequenced using primers based on the known cDNA sequence. This step may be repeated until a full
5 length cDNA is obtained.

One method for identifying all 3' isoforms is to PCR amplify the 3' ends of the variant cDNA from a variety of tissues including but not limiting to, choroid plexus, hypothalamus, fetal liver, bone marrow, ovary, or prostate.
10 To obtain the 3' end of the cDNA, an oligo-dT primer is used to synthesize the cDNA first strand. OB-R specific primers from the conserved region of the gene (e.g. up stream of nucleotide 2770) and oligo-dT are then used to amplify the 3' end. The PCR fragments are cloned and sequenced by standard
15 techniques. Once obtained, these sequences may be translated into amino acid sequence and examined for certain landmarks such as continuous open reading frame, regulatory regions that associate with tyrosine kinase activation, and finally overall structural similarity to known OB-R variants.

20

5.2. EXPRESSION OF THE OB-R VARIANT

In accordance with the invention, the OB-R variant polynucleotide sequence which encodes a protein, peptide fragments, fusion proteins or functional equivalents thereof,
25 may be used to generate recombinant DNA molecules that direct the expression of the protein, peptide fragments, fusion proteins or a functional equivalent thereof, in appropriate host cells. Such polynucleotide sequences, as well as other polynucleotides which selectively hybridize to at least a
30 part of such polynucleotides or their complements, may also be used in nucleic acid hybridization assays, Southern and Northern blot analyses, etc.

Due to the inherent degeneracy of the genetic code, other DNA sequences which encode substantially the same or a
35 functionally equivalent amino acid sequence, may be used in the practice of the invention for the expression of the OB-R variant. Such DNA sequences include those which are capable

of hybridizing to the OB-R variant sequence under stringent conditions, particularly at its 3' end. The phrase "stringent conditions" as used herein refers to those hybridizing conditions that (1) employ low ionic strength and high temperature for washing, for example, 0.015 M NaCl/0.0015 M sodium citrate/0.1% SDS at 50°C.; (2) employ during hybridization a denaturing agent such as formamide, for example, 50% (vol/vol) formamide with 0.1% bovine serum albumin/0.1% Ficoll/0.1% polyvinylpyrrolidone/50 mM sodium phosphate buffer at pH 6.5 with 750 mM NaCl, 75 mM sodium citrate at 42°C; or (3) employ 50% formamide, 5 x SSC (0.75 M NaCl, 0.075 M Sodium pyrophosphate, 5 x Denhardt's solution, sonicated salmon sperm DNA (50 g/ml), 0.1% SDS, and 10% dextran sulfate at 42°C, with washes at 42°C in 0.2 x SSC and 0.1% SDS.

Altered DNA sequences which may be used in accordance with the invention include deletions, additions or substitutions of different nucleotide residues resulting in a sequence that encodes the same or a functionally equivalent gene product. The gene product itself may contain deletions, additions or substitutions of amino acid residues within the OB-R variant sequence, which result in a silent change thus producing a functionally equivalent protein. Such amino acid substitutions may be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues involved. For example, negatively charged amino acids include aspartic acid and glutamic acid; positively charged amino acids include lysine, histidine and arginine; amino acids with uncharged polar head groups having similar hydrophilicity values include the following: glycine, asparagine, glutamine, serine, threonine, tyrosine; and amino acids with nonpolar head groups include alanine, valine, isoleucine, leucine, phenylalanine, proline, methionine, tryptophan.

The DNA sequence of the invention may be engineered in order to alter the OB-R variant coding sequence for a variety of ends, including but not limited to, alterations which

modify processing and expression of the gene product. For example, mutations may be introduced using techniques which are well known in the art, e.g., site-directed mutagenesis, to insert new restriction sites, to alter glycosylation patterns, phosphorylation, etc. In addition, the intracellular domain may also be altered and replaced by a different domain, such as the OB-R intracellular domain by Tartaglia et al.

In another embodiment of the invention, the OB-R variant sequence may be ligated to a heterologous sequence to encode a fusion protein. For example, for screening of peptide libraries for inhibitors or stimulators of receptor activity, it may be useful to encode a chimeric protein expressing a heterologous epitope that is recognized by a commercially available antibody. A fusion protein may also be engineered to contain a cleavage site located between the OB-R variant sequence and the heterologous protein sequence, so that the variant may be cleaved away from the heterologous moiety.

In an alternate embodiment of the invention, the coding sequence of the OB-R variant could be synthesized in whole or in part, using chemical methods well known in the art. (See, for example, Caruthers et al., 1980, *Nuc. Acids Res. Symp. Ser.* 7:215-233; Crea and Horn, 180, *Nuc. Acids Res.* 9(10):2331; Matteucci and Caruthers, 1980, *Tetrahedron Letters* 21:719; and Chow and Kempe, 1981, *Nuc. Acids Res.* 9(12):2807-2817). Alternatively, the protein itself could be produced using chemical methods to synthesize OB-R variant amino acid sequence in whole or in part. For example, peptides can be synthesized by solid phase techniques, cleaved from the resin, and purified by preparative high performance liquid chromatography. (e.g., see Creighton, 1983, *Proteins Structures And Molecular Principles*, W.H. Freeman and Co., N.Y. pp. 50-60). The composition of the synthetic peptides may be confirmed by amino acid analysis or sequencing (e.g., the Edman degradation procedure; see Creighton, 1983, *Proteins, Structures and Molecular Principles*, W.H. Freeman and Co., N.Y., pp. 34-49).

In order to express the OB-R variant in host cells, the nucleotide sequence coding for the variant, or a functional equivalent, is inserted into an appropriate expression vector, i.e., a vector which contains the necessary elements for the transcription and translation of the inserted coding sequence. The expressed gene products as well as host cells or cell lines transfected or transformed with recombinant OB-R variant expression vectors can be used for a variety of purposes. For example, host cells expressing the OB-R variant may be used to verify the ability of this molecule to bind leptin in a binding assay with radiolabeled, enzyme-conjugated or fluorescent dye-conjugated leptin. At the same time, the ability of the molecule to transduce an activation signal in host cells upon binding to leptin may be tested by assaying proliferation or phosphorylation pattern of kinases in the cells. In addition, genetically-engineered host cells can be used to screen for and select agonist and antagonist compounds, including any inhibitors that would interfere with binding of leptin to the extracellular domain of the OB-R variant. In that connection, such host cells may be used to screen for and select small molecules that can supplement the incomplete signal transduced by the OB-R variant following leptin binding. Other uses, include, but are not limited to generating antibodies (i.e., monoclonal or polyclonal) that competitively inhibit activity of an OB-R variant, neutralize its activity, or even enhances its activity. Antibodies may be used in detecting and quantifying expression of OB-R levels in cells and tissues.

5.3. USES OF THE OB-R VARIANT POLYNUCLEOTIDE

The OB-R variant polynucleotide may be used for diagnostic and/or therapeutic purposes. For diagnostic purposes, the OB-R variant polynucleotide may be used to detect gene expression or aberrant gene expression in obese individuals as well as in normal individuals to identify predisposition for obesity. Included in the scope of the invention are oligonucleotide sequences, that include

antisense RNA and DNA molecules, ribozymes and triplex DNA, that function to inhibit translation of OB-R variant.

5.3.1. DIAGNOSTIC USES OF OB-R VARIANT POLYNUCLEOTIDE

5 The OB-R variant polynucleotide may have a number of uses for the diagnosis of the possible causes underlying obesity, resulting from expression of the receptor variant. For example, the OB-R variant cytoplasmic domain DNA sequence may be used in hybridization assays of biopsies or autopsies
10 to diagnose OB-R variant expression; e.g., Southern or Northern analysis, including *in situ* hybridization assays as well as PCR. Such techniques are well known in the art, and are in fact the basis of many commercially available diagnostic kits. For PCR detection, primers may be designed
15 from a conserved region of the coding sequence and within the 3' region of OB-R variant. The tissues suitable for such analysis include but are not limited to, hypothalamus, choroid plexus, adipose tissues, lung, prostate, ovary, small intestine, bone marrow and peripheral blood mononuclear
20 cells.

5.3.2. THERAPEUTIC USES OF THE OB-R VARIANT POLYNUCLEOTIDE

The OB-R variant polynucleotide may be useful in the
25 treatment of various abnormal obese conditions. By introducing gene sequences into cells, gene therapy can be used to treat conditions in which the cells do not respond to leptin normally due to expression of the OB-R variant. In some instances, the polynucleotide encoding a functional OB-R
30 is intended to replace or act in the place of the functionally deficient OB-R variant gene. Alternatively, abnormal conditions characterized by expression of two copies of the OB-R variant can be treated using the gene therapy techniques described below.

35 Non-responsiveness to normal levels of leptin is an important cause of obesity. This may result from a functionally defective receptor that does not transduce

competent signals upon ligand binding. Recombinant gene therapy vectors, such as viral vectors, may be engineered to express signalling competent forms of OB-R which may be used to augment the non-responsiveness of the naturally occurring OB-R variant. A signalling competent form may be, for example, a protein with the same extracellular domain and transmembrane region, but containing all or part of its normal signal transduction domain, such as that described by Tartaglia et al. (1995, *Cell* 83:1263-1271). Thus recombinant gene therapy vectors may be used therapeutically for treatment of obesity resulting from expression or activity of the OB-R variant. Accordingly, the invention provides a method of augmenting signal transduction by an endogenous OB-R variant in a cell comprising delivering a DNA molecule encoding a signalling competent form of the OB-R to the cell so that the signalling competent protein is produced in the cell and competes with the endogenous defective OB-R variant for access to molecules in the signalling pathway which does not activate or are not activated by the endogenous natural defective receptor. Additionally, since dimerization of a functional receptor with a defective variant may occur in cells of heterozygous individuals, small molecules may be used to inhibit such pairing, thereby increasing the number of functional dimeric receptors for proper signalling in response to leptin.

In contrast, overexpression of either leptin or a competent OB-R may result in a clinical anorexic-like syndrome due to a loss of appetite or hypermetabolic activity. In such cases, the OB-R variant of the invention may be introduced into cells with functional receptors to cause a decrease in the number of functional receptors or to compete with such receptors for leptin binding.

Expression vectors derived from viruses such as retroviruses, vaccinia virus, adeno-associated virus, herpes viruses, or bovine papilloma virus, may be used for delivery of recombinant functional OB-R into the targeted cell population. Methods which are well known to those skilled in

the art can be used to construct recombinant viral vectors containing an OB-R polynucleotide sequence. See, for example, the techniques described in Sambrook et al., 1989, Molecular Cloning A Laboratory Manual, Cold Spring Harbor Laboratory, N.Y. and Ausubel et al., 1989, Current Protocols in Molecular Biology, Greene Publishing Associates and Wiley Interscience, N.Y. Alternatively, recombinant OB-R molecules can be reconstituted into liposomes for delivery to target cells.

- 10 Oligonucleotide sequences including anti-sense RNA and DNA molecules and ribozymes that function to inhibit the translation of the OB-R variant mRNA are within the scope of the invention. Anti-sense RNA and DNA molecules act to directly block the translation of mRNA by binding to targeted
15 mRNA and preventing protein translation. In regard to antisense DNA, oligodeoxyribonucleotides derived from the OB-R variant nucleotide sequence at nucleotide #2771 and beyond, are preferred.

- Ribozymes are enzymatic RNA molecules capable of
20 catalyzing the specific cleavage of RNA. The mechanism of ribozyme action involves sequence specific hybridization of the ribozyme molecule to complementary target RNA, followed by endonucleolytic cleavage. Within the scope of the invention are engineered hammerhead motif ribozyme molecules
25 that specifically and efficiently catalyze endonucleolytic cleavage of OB-R variant RNA sequences.

- Specific ribozyme cleavage sites within any potential RNA target are initially identified by scanning the target molecule for ribozyme cleavage sites which include the
30 following sequences, GUA, GUU and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides corresponding to the region of the target gene containing the cleavage site may be evaluated for predicted structural features such as secondary structure that may render the
35 oligonucleotide sequence unsuitable. The suitability of candidate targets may also be evaluated by testing their

accessibility to hybridization with complementary oligonucleotides, using ribonuclease protection assays.

Oligodeoxyribonucleotides can form sequence-specific triple helices by hydrogen bonding to specific complementary sequences in duplexed DNA. Interest in triple helices has focused on the potential biological and therapeutic applications of these structures. Formation of specific triple helices may selectively inhibit the replication and/or gene expression of targeted genes by prohibiting the specific binding of functional trans-acting factors.

Oligonucleotides to be used in triplex helix formation should be single stranded and composed of deoxynucleotides. The base composition of these oligonucleotides must be designed to promote triple helix formation via Hoogsteen base pairing rules, which generally require sizeable stretches of either purines or pyrimidines to be present on one strand of a duplex. Oligonucleotide sequences may be pyrimidine-based, which will result in TAT and CGC triplets across the three associated strands of the resulting triple helix. The pyrimidine-rich oligonucleotides provide base complementarity to a purine-rich region of a single strand of the duplex in a parallel orientation to that strand. In addition, oligonucleotides may be chosen that are purine-rich, for example, containing a stretch of G residues. These oligonucleotides will form a triple helix with a DNA duplex that is rich in GC pairs, in which the majority of the purine residues are located on a single strand of the targeted duplex, resulting in GGC triplets across the three strands in the triplex. Alternatively, the potential sequences that can be targeted for triple helix formation may be increased by creating a so called "switchback" oligonucleotide. Switchback oligonucleotides are synthesized in an alternating 5'-3', 3'-5' manner, such that they base pair with first one strand of a duplex and then the other, eliminating the necessity for a sizeable stretch of either purines or pyrimidines to be present on one strand of a duplex.

Both anti-sense RNA and DNA molecules and ribozymes of the invention may be prepared by any method known in the art for the synthesis of RNA molecules. These include techniques for chemically synthesizing oligodeoxyribonucleotides well known in the art such as solid phase phosphoramidite chemical synthesis. Alternatively, RNA molecules may be generated by *in vitro* and *in vivo* transcription of DNA sequences encoding the antisense RNA molecule. Such DNA sequences may be incorporated into a wide variety of vectors which incorporate suitable RNA polymerase promoters such as the T7 or SP6 polymerase promoters. Alternatively, antisense cDNA constructs that synthesize antisense RNA constitutively or inducibly, depending on the promoter used, can be introduced stably into cell lines.

Various modifications to the DNA molecules may be introduced as a means of increasing intracellular stability and half-life. Possible modifications include but are not limited to the addition of flanking sequences of ribo- or deoxy- nucleotides to the 5' and/or 3' ends of the molecule or the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages within the oligodeoxyribonucleotide backbone.

5.4. ACTIVATION OF TYROSINE KINASE PATHWAYS IN OBESITY

Many known class I cytokine receptors initiate cell signaling via Janus kinases (JAKs) (Ihle, 1995, *Nature* 377:591-594; Heldin, 1995, *Cell* 80:213-223; Kishimoto et al, 1994, *Cell* 76:253-62; Ziemiecki et al, 1994, *Trends Cell. Biol.* 4:207-212). JAK1-3 have been shown to bind to conserved sequences termed box1 and box2 (Fukunaga et al., 1991, *EMBO J.* 10:2855-65; Murakami, 1991, *Proc. Natl. Acad. Sci. USA* 88:11349-53). Ligand binding induces a homo- or hetero-dimerization of receptor chains which activates, by phosphorylation, the JAKs. The activated JAKs, in turn, phosphorylate members of the STAT family (Heldin, 1995, *Cell* 80:213-223; Kishimoto et al., *Blood* 86:1243-54; Darnell et al., 1994, *Science* 264:1415-21; Zhong et al, 1994, *Proc.*

Natl. Acad. Sci. USA 91:4806-10; Hou et al., 1994, Science 265:1701-6). These phosphorylated STATs ultimately translocate to the nucleus, form transcription complexes, and regulate gene expression. Both box1 and box2 are required for complete signaling in certain systems. (Fukunaga et al., 1991, EMBO J. 10:2855-65; Murakami, 1991, Proc. Natl. Acad. Sci. USA 88:11349-53). The OB-R variant disclosed herein has a typical box1 (from nucleotide #2707-2730) that contains the critical xWxxxPxP amino acid sequence, but it does not contain an obvious box2 nor further downstream sequences that are important for normal receptor activation. Therefore, it is possible to use compounds that activate JAKs to directly activate these pathways for weight regulation without triggering the OB-R.

15

6. EXAMPLE: MOLECULAR CLONING OF AN OB-R VARIANT

A number of cDNA clones were isolated from a human fetal liver cDNA library (Clontech, Palo Alto, CA), and the DNA sequences of several of these clones were determined. These clones (designated as Hu-B1.219 #4, #33, #34, #1, #8) contained overlapping sequences, which were then compiled into a contiguous nucleotide sequence (Figure 1A-1E). When the deduced amino acid sequence of one such sequence was compared with the sequence of a recently published human OB-R, they were shown to be nearly identical in the extracellular domains with the exception of three amino acids, whereas there existed extensive diversity in their intracellular cytoplasmic domains at the 3' end. The predicted protein sequence contains two FN III domains, each containing a "WS box", which are characteristic of genes of the Class I cytokine receptor family. Therefore, the cDNA disclosed herein encodes an OB-R variant.

When various human tissue RNA were probed with a fragment of this OB-R variant by Northern blot analysis, expression of this molecule was detected in heart, placenta, lung, liver, muscle, pancreas, prostate, ovary, small intestine and brain.

Based on the sequence presented in Figure 1A-1E, the translation initiation site appears at position #97. The sequence encodes an open reading frame up to and including nucleotide #2814. It is believed that the sequence between 5 nucleotides #2629 and #2682 encodes a transmembrane domain. The complete sequence encodes a protein of 906 amino acids.

The sequence of the OB-R variant is identical to the sequence of human OB-R reported by Tartaglia (1995, Cell 83:1263-1271) in the transmembrane region and a portion of 10 the intracellular domain up to and including nucleotide #2769, then they diverge at nucleotide #2770 and beyond. In addition, the product of this cDNA is substantially shorter in its intracellular domain than the published human OB-R. These two forms of OB-R may derive from a common precursor 15 mRNA by an alternative splicing mechanism. The sequence in this region is consistent with well known splice junctions.

7. DEPOSIT OF MICROORGANISMS

The following organisms were deposited with the American 20 Type Culture Collection (ATCC), 12301 Parklawn Drive, Rockville, Maryland 20852.

	<u>Strain Designation</u>	<u>Accession No.</u>
--	---------------------------	----------------------

	HuB1.219, #1	75885
	HuB1.219, #4	75886
25	HuB1.219, #33	75888
	HuB1.219, #34	75889
	HuB1.219, #8	75974

The present invention is not to be limited in scope by the exemplified embodiments, which are intended as 30 illustrations of individual aspects of the invention.

Indeed, various modifications for the invention in addition to those shown and described herein will become apparent to those skilled in the art from the foregoing description and accompanying drawings. Such modifications are intended to 35 fall within the scope of the appended claims.

All publications cited herein are incorporated by reference in their entirety.

(2) GENERAL INFORMATION:

- (2) INFORMATION FOR SEQ ID NO:1:

- (ii) MOLECULE TYPE: DNA

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

GCG CGC GCG ACG CAG GTG CCC GAG CCC CGG CCC GCG CCC ATC TCT GCC
 Ala Arg Ala Thr Gln Val Pro Glu Pro Arg Pro Ala Pro Ile Ser Ala 48
 1 5 10 15
 TTC GGT CGA GTT GGA CCC CCG GAT CAA GGT GTA CTT CTC TGA AGT AAG 96

Phe Gly Arg Val Gly Pro Pro Asp Gln Gly Val Leu Leu * Ser Lys	
20 25 30	
ATG ATT TGT CAA AAA TTC TGT GTG GTT TTG TTA CAT TGG GAA TTT ATT	144
Met Ile Cys Gln Lys Phe Cys Val Val Leu Leu His Trp Glu Phe Ile	
35 40 45	
TAT GTG ATA ACT GCG TTT AAC TTG TCA TAT CCA ATT ACT CCT TGG AGA	192
Tyr Val Ile Thr Ala Phe Asn Leu Ser Tyr Pro Ile Thr Pro Trp Arg	
50 55 60	
TTT AAG TTG TCT TGC ATG CCA CCA AAT TCA ACC TAT GAC TAC TTC CTT	240
Phe Lys Leu Ser Cys Met Pro Pro Asn Ser Thr Tyr Asp Tyr Phe Leu	
65 70 75 80	
TTG CCT GCT GGA CTC TCA AAG AAT ACT TCA AAT TCG AAT GGA CAT TAT	288
Leu Pro Ala Gly Leu Ser Lys Asn Thr Ser Asn Ser Asn Gly His Tyr	
85 90 95	
GAG ACA GCT GTT GAA CCT AAG TTT AAT TCA AGT GGT ACT CAC TTT TCT	336
Glu Thr Ala Val Glu Pro Lys Phe Asn Ser Ser Gly Thr His Phe Ser	
100 105 110	
AAC TTA TCC AAA GCA ACT TTC CAC TGT TGC TTT CGG AGT GAG CAA GAT	384
Asn Leu Ser Lys Ala Thr Phe His Cys Cys Phe Arg Ser Glu Gln Asp	
115 120 125	
AGA AAC TGC TCC TTA TGT GCA GAC AAC ATT GAA GGA AGG ACA TTT GTT	432
Arg Asn Cys Ser Leu Cys Ala Asp Asn Ile Glu Gly Arg Thr Phe Val	
130 135 140	
TCA ACA GTA AAT TCT TTA GTT TTT CAA CAA ATA GAT GCA AAC TGG AAC	480
Ser Thr Val Asn Ser Leu Val Phe Gln Gln Ile Asp Ala Asn Trp Asn	
145 150 155 160	
ATA CAG TGC TGG CTA AAA GGA GAC TTA AAA TTT CTC ATC TGT TAT GTG	528
Ile Gln Cys Trp Leu Lys Gly Asp Leu Lys Leu Phe Ile Cys Tyr Val	
165 170 175	
GAG TCA TTA TTT AAG AAT CTA TTC AGG AAT TAT AAC TAT AAG GTC CAT	576
Glu Ser Leu Phe Lys Asn Leu Phe Arg Asn Tyr Asn Tyr Lys Val His	
180 185 190	
CTT TTA TAT GTT CTG CCT GAA GTG TTA GAA GAT TCA CCT CTG GTT CCC	624
Leu Leu Tyr Val Leu Pro Glu Val Leu Glu Asp Ser Pro Leu Val Pro	
195 200 205	
CAA AAA GGC AGT TTT CAG ATG GTT CAC TGC AAT TGC AGT GTT CAT GAA	672
Gln Lys Gly Ser Phe Gln Met Val His Cys Asn Cys Ser Val His Glu	
210 215 220	
TGT TGT GAA TGT CTT GTG CCT GTG CCA ACA GCC AAA CTC AAC GAC ACT	720
Cys Cys Glu Cys Leu Val Pro Val Pro Thr Ala Lys Leu Asn Asp Thr	
225 230 235 240	
CTC CTT ATG TGT TTG AAA ATC ACA TCT GGT GGA GTA ATT TTC CGG TCA	768
Leu Leu Met Cys Leu Lys Ile Thr Ser Gly Gly Val Ile Phe Arg Ser	
245 250 255	
CCT CTA ATG TCA GTT CAG CCA ATA AAT ATG GTG AAG CCT GAT CCA CCA	816
Pro Leu Met Ser Val Gln Pro Ile Asn Met Val Lys Pro Asp Pro Pro	
260 265 270	
TTA GGT TTG CAT ATG GAA ATC ACA GAT GAT GGT AAT TTA AAG ATT TCT	864
Leu Gly Leu His Met Glu Ile Thr Asp Asp Gly Asn Leu Lys Ile Ser	
275 280 285	

TGG TCC AGC CCA-CCA TTG GTA CCA TTT CCA CTT CAA TAT CAA GTG AAA Trp Ser Ser Pro Pro Leu Val Pro Phe Pro Leu Gln Tyr Gln Val Lys 290 295 300	912
TAT TCA GAG AAT TCT ACA ACA GTT ATC AGA GAA GCT GAC AAG ATT GTC Tyr Ser Glu Asn Ser Thr Thr Val Ile Arg Glu Ala Asp Lys Ile Val 305 310 315 320	960
TCA GCT ACA TCC CTG CTA GTA GAC AGT ATA CTT CCT GGG TCT TCG TAT Ser Ala Thr Ser Leu Leu Val Asp Ser Ile Leu Pro Gly Ser Ser Tyr 325 330 335	1008
GAG GTT CAG GTG AGG GGC AAG AGA CTG GAT GGC CCA GGA ATC TGG AGT Glu Val Gln Val Arg Gly Lys Arg Leu Asp Gly Pro Gly Ile Trp Ser 340 345 350	1056
GAC TGG AGT ACT CCT CGT GTC TTT ACC ACA CAA GAT GTC ATA TAC TTT Asp Trp Ser Thr Pro Arg Val Phe Thr Thr Gln Asp Val Ile Tyr Phe 355 360 365	1104
CCA CCT AAA ATT CTG ACA AGT GTT GGG TCT AAT GTT TCT TTT CAC TGC Pro Pro Lys Ile Leu Thr Ser Val Gly Ser Asn Val Ser Phe His Cys 370 375 380	1152
ATC TAT AAG AAG GAA AAC AAG ATT GTT CCC TCA AAA GAG ATT GTT TGG Ile Tyr Lys Lys Glu Asn Lys Ile Val Pro Ser Lys Glu Ile Val Trp 385 390 395 400	1200
TGG ATG AAT TTA GCT GAG AAA ATT CCT CAA AGC CAG TAT GAT GTT GTG Trp Met Asn Leu Ala Glu Lys Ile Pro Gln Ser Gln Tyr Asp Val Val 405 410 415	1248
AGT GAT CAT GTT AGC AAA GTT ACT TTT TTC AAT CTG AAT GAA ACC AAA Ser Asp His Val Ser Lys Val Thr Phe Asn Leu Asn Glu Thr Lys 420 425 430	1296
CCT CGA GGA AAG TTT ACC TAT GAT GCA GTG TAC TGC TGC AAT GAA CAT Pro Arg Gly Lys Phe Thr Tyr Asp Ala Val Tyr Cys Cys Asn Glu His 435 440 445	1344
GAA TGC CAT CAT CGC TAT GCT GAA TTA TAT GTG ATT GAT GTC AAT ATC Glu Cys His His Arg Tyr Ala Glu Leu Tyr Val Ile Asp Val Asn Ile 450 455 460	1392
AAT ATC TCA TGT GAA ACT GAT GGG TAC TTA ACT AAA ATG ACT TGC AGA Asn Ile Ser Cys Glu Thr Asp Gly Tyr Leu Thr Lys Met Thr Cys Arg 465 470 475 480	1440
TGG TCA ACC AGT ACA ATC CAG TCA CTT GCG GAA AGC ACT TTG CAA TTG Trp Ser Thr Ser Thr Ile Gln Ser Leu Ala Glu Ser Thr Leu Gln Leu 485 490 495	1488
AGG TAT CAT AGG AGC AGC CTT TAC TGT TCT GAT ATT CCA TCT ATT CAT Arg Tyr His Arg Ser Ser Leu Tyr Cys Ser Asp Ile Pro Ser Ile His 500 505 510	1536
CCC ATA TCT GAG CCC AAA GAT TGC TAT TTG CAG AGT GAT GGT TTT TAT Pro Ile Ser Glu Pro Lys Asp Cys Tyr Leu Gln Ser Asp Gly Phe Tyr 515 520 525	1584
GAA TGC ATT TTC CAG CCA ATC TTC CTA TTA TCT GGC TAC ACA ATG TGG Glu Cys Ile Phe Gln Pro Ile Phe Leu Leu Ser Gly Tyr Thr Met Trp 530 535 540	1632
ATT AGG ATC AAT CAC TCT CTA GGT TCA CTT GAC TCT CCA CCA ACA TGT Ile Arg Ile Asn His Ser Leu Gly Ser Leu Asp Ser Pro Pro Thr Cys 545 550 555 560	1680

GTC	CTT	CCT	GAT	TCT	GTG	GTG	AAG	CCA	CTG	CCT	CCA	TCC	AGT	GTG	AAA	1728
Val	Leu	Pro	Asp	Ser	Val	Val	Lys	Pro	Leu	Pro	Pro	Ser	Ser	Val	Lys	
				565					570					575		
CGA	GAA	ATT	ACT	ATA	AAC	ATT	GGA	TTA	TTG	AAA	ATA	TCT	TGG	GAA	AAG	1776
Arg	Glu	Ile	Thr	Ile	Asn	Ile	Gly	Leu	Leu	Lys	Ile	Ser	Trp	Glu	Lys	
			580					585					590			
CCA	GTC	TTT	CCA	GAG	AAT	AAC	CTT	CAA	TTC	CAG	ATT	CGC	TAT	GGT	TTA	1824
Pro	Val	Phe	Pro	Glu	Asn	Asn	Leu	Gln	Phe	Gln	Ile	Arg	Tyr	Gly	Leu	
		595					600					605				
AGT	GGA	AAA	GAA	GTA	CAA	TGG	AAG	ATG	TAT	GAG	GTT	TAT	GAT	CGA	AAA	1872
Ser	Gly	Lys	Glu	Val	Gln	Trp	Lys	Met	Tyr	Glu	Val	Tyr	Asp	Arg	Lys	
	610					615					620					
TCA	AAA	TCT	GTC	AGT	CTC	CCA	GTT	CCA	GAC	TTG	TGT	GCA	GTC	TAT	GCT	1920
Ser	Lys	Ser	Val	Ser	Leu	Pro	Val	Pro	Asp	Leu	Cys	Ala	Val	Tyr	Ala	
	625				630				635						640	
GTT	CAG	GTG	CGC	TGT	AAG	AGG	CTA	GAT	GGA	CTG	GGA	TAT	TGG	AGT	AAT	1968
Val	Gln	Val	Arg	Cys	Lys	Arg	Leu	Asp	Gly	Leu	Gly	Tyr	Trp	Ser	Asn	
				645					650					655		
TGG	AGC	AAT	CCA	GCC	TAC	ACA	GTT	GTC	ATG	GAT	ATA	AAA	GTT	CCT	ATG	2016
Trp	Ser	Asn	Pro	Ala	Tyr	Thr	Val	Val	Met	Asp	Ile	Lys	Val	Pro	Met	
			660					665					670			
AGA	GGA	CCT	GAA	TTT	TGG	AGA	ATA	ATT	AAT	GGA	GAT	ACT	ATG	AAA	AAG	2064
Arg	Gly	Pro	Glu	Phe	Trp	Arg	Ile	Ile	Asn	Gly	Asp	Thr	Met	Lys	Lys	
		675					680					685				
GAG	AAA	AAT	GTC	ACT	TTA	CTT	TGG	AAG	CCC	CTG	ATG	AAA	AAT	GAC	TCA	2112
Glu	Lys	Asn	Val	Thr	Leu	Leu	Trp	Lys	Pro	Leu	Met	Lys	Asn	Asp	Ser	
	690					695					700					
TTG	TGC	AGT	GTT	CAG	AGA	TAT	GTG	ATA	AAC	CAT	CAT	ACT	TCC	TGC	AAT	2160
Leu	Cys	Ser	Val	Gln	Arg	Tyr	Val	Ile	Asn	His	His	Thr	Ser	Cys	Asn	
	705				710					715					720	
GGA	ACA	TGG	TCA	GAA	GAT	GTG	GGA	AAT	CAC	ACG	AAA	TTC	ACT	TTC	CTG	2208
Gly	Thr	Trp	Ser	Glu	Asp	Val	Gly	Asn	His	Thr	Lys	Phe	Thr	Phe	Leu	
			725						730					735		
TGG	ACA	GAG	CAA	GCA	CAT	ACT	GTT	ACG	GTT	CTG	GCC	ATC	AAT	TCA	ATT	2256
Trp	Thr	Glu	Gln	Ala	His	Thr	Val	Thr	Val	Leu	Ala	Ile	Asn	Ser	Ile	
			740					745					750			
GGT	GCT	TCT	GTT	GCA	AAT	TTT	AAT	TTA	ACC	TTT	TCA	TGG	CCT	ATG	AGC	2304
Gly	Ala	Ser	Val	Ala	Asn	Phe	Asn	Leu	Thr	Phe	Ser	Trp	Pro	Met	Ser	
		755					760					765				
AAA	GTA	AAT	ATC	GTG	CAG	TCA	CTC	AGT	GCT	TAT	CCT	TTA	AAC	AGC	AGT	2352
Lys	Val	Asn	Ile	Val	Gln	Ser	Leu	Ser	Ala	Tyr	Pro	Leu	Asn	Ser	Ser	
	770					775					780					
TGT	GTG	ATT	GTT	TCC	TGG	ATA	CTA	TCA	CCC	AGT	GAT	TAC	AAG	CTA	ATG	2400
Cys	Val	Ile	Val	Ser	Trp	Ile	Leu	Ser	Pro	Ser	Asp	Tyr	Lys	Leu	Met	
	785					790				795					800	
TAT	TTT	ATT	ATT	GAG	TGG	AAA	AAT	CTT	AAT	GAA	GAT	GGT	GAA	ATA	AAA	2448
Tyr	Phe	Ile	Ile	Glu	Trp	Lys	Asn	Leu	Asn	Glu	Asp	Gly	Glu	Ile	Lys	
				805					810					815		
TGG	CTT	AGA	ATC	TCT	TCA	TCT	GTT	AAG	AAG	TAT	TAT	ATC	CAT	GAT	CAT	2496
Trp	Leu	Arg	Ile	Ser	Ser	Ser	Val	Lys	Lys	Tyr	Tyr	Ile	His	Asp	His	
			820					825					830			

TTT ATC CCC ATT GAG AAG TAC CAG TTC AGT CTT TAC CCA ATA TTT ATG Phe Ile Pro Ile Glu Lys Tyr Gln Phe Ser Leu Tyr Pro Ile Phe Met 835 840 845	2544
GAA GGA GTG GGA AAA CCA AAG ATA ATT AAT AGT TTC ACT CAA GAT GAT Glu Gly Val Gly Lys Pro Lys Ile Ile Asn Ser Phe Thr Gln Asp Asp 850 855 860	2592
ATT GAA AAA CAC CAG AGT GAT GCA GGT TTA TAT GTA ATT GTG CCA GTA Ile Glu Lys His Gln Ser Asp Ala Gly Leu Tyr Val Ile Val Pro Val 865 870 875 880	2640
ATT ATT TCC TCT TCC ATC TTA TTG CTT GGA ACA TTA TTA ATA TCA CAC Ile Ile Ser Ser Ser Ile Leu Leu Leu Gly Thr Leu Leu Ile Ser His 885 890 895	2688
CAA AGA ATG AAA AAG CTA TTT TGG GAA GAT GTT CCG AAC CCC AAG AAT Gln Arg Met Lys Lys Leu Phe Trp Glu Asp Val Pro Asn Pro Lys Asn 900 905 910	2736
TGT TCC TGG GCA CAA GGA CCT AAT TTT CAG AAG AAA ATG CCT GGC ACA Cys Ser Trp Ala Gln Gly Pro Asn Phe Gln Lys Lys Met Pro Gly Thr 915 920 925	2784
AAG GAA CTA CTG GGT GGA GGT TGG TTG ACT TAG GAA ATG CTT GTG AAG Lys Glu Leu Leu Gly Gly Gly Trp Leu Thr * Glu Met Leu Val Lys 930 935 940	2832
CTA CGT CCT ACC TCG TGC GCA CCT GCT CTC CCT GAG GTG TGC ACA ATG Leu Arg Pro Thr Ser Cys Ala Pro Ala Leu Pro Glu Val Cys Thr Met 945 950 955 960	2880

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 29 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Ala Arg Ala Thr Gln Val Pro Glu Pro Arg Pro Ala Pro Ile Ser Ala
1 5 10 15
Phe Gly Arg Val Gly Pro Pro Asp Gln Gly Val Leu Leu
20 25

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 908 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Ser Lys Met Ile Cys Gln Lys Phe Cys Val Val Leu Leu His Trp Glu
1 5 10 15
Phe Ile Tyr Val Ile Thr Ala Phe Asn Leu Ser Tyr Pro Ile Thr Pro

22

Val Trp Trp Met Asn Leu Ala Glu Lys Ile Pro Gln Ser Gln Tyr Asp
 370 375 380
 Val Val Ser Asp His Val Ser Lys Val Thr Phe Phe Asn Leu Asn Glu
 385 390 395 400
 Thr Lys Pro Arg Gly Lys Phe Thr Tyr Asp Ala Val Tyr Cys Cys Asn
 405 410 415
 Glu His Glu Cys His His Arg Tyr Ala Glu Leu Tyr Val Ile Asp Val
 420 425 430
 Asn Ile Asn Ile Ser Cys Glu Thr Asp Gly Tyr Leu Thr Lys Met Thr
 435 440 445
 Cys Arg Trp Ser Thr Ser Thr Ile Gln Ser Leu Ala Glu Ser Thr Leu
 450 455 460
 Gln Leu Arg Tyr His Arg Ser Ser Leu Tyr Cys Ser Asp Ile Pro Ser
 465 470 475 480
 Ile His Pro Ile Ser Glu Pro Lys Asp Cys Tyr Leu Gln Ser Asp Gly
 485 490 495
 Phe Tyr Glu Cys Ile Phe Gln Pro Ile Phe Leu Leu Ser Gly Tyr Thr
 500 505 510
 Met Trp Ile Arg Ile Asn His Ser Leu Gly Ser Leu Asp Ser Pro Pro
 515 520 525
 Thr Cys Val Leu Pro Asp Ser Val Val Lys Pro Leu Pro Pro Ser Ser
 530 535 540
 Val Lys Arg Glu Ile Thr Ile Asn Ile Gly Leu Leu Lys Ile Ser Trp
 545 550 555 560
 Glu Lys Pro Val Phe Pro Glu Asn Asn Leu Gln Phe Gln Ile Arg Tyr
 565 570 575
 Gly Leu Ser Gly Lys Glu Val Gln Trp Lys Met Tyr Glu Val Tyr Asp
 580 585 590
 Arg Lys Ser Lys Ser Val Ser Leu Pro Val Pro Asp Leu Cys Ala Val
 595 600 605
 Tyr Ala Val Gln Val Arg Cys Lys Arg Leu Asp Gly Leu Gly Tyr Trp
 610 615 620
 Ser Asn Trp Ser Asn Pro Ala Tyr Thr Val Val Met Asp Ile Lys Val
 625 630 635 640
 Pro Met Arg Gly Pro Glu Phe Trp Arg Ile Ile Asn Gly Asp Thr Met
 645 650 655
 Lys Lys Glu Lys Asn Val Thr Leu Leu Trp Lys Pro Leu Met Lys Asn
 660 665 670
 Asp Ser Leu Cys Ser Val Gln Arg Tyr Val Ile Asn His His Thr Ser
 675 680 685
 Cys Asn Gly Thr Trp Ser Glu Asp Val Gly Asn His Thr Lys Phe Thr
 690 695 700
 Phe Leu Trp Thr Glu Gln Ala His Thr Val Thr Val Leu Ala Ile Asn
 705 710 715 720
 Ser Ile Gly Ala Ser Val Ala Asn Phe Asn Leu Thr Phe Ser Trp Pro

725	730	735
Met Ser Lys Val Asn Ile Val Gln Ser Leu Ser Ala Tyr Pro Leu Asn 740 745 750		
Ser Ser Cys Val Ile Val Ser Trp Ile Leu Ser Pro Ser Asp Tyr Lys 755 760 765		
Leu Met Tyr Phe Ile Ile Glu Trp Lys Asn Leu Asn Glu Asp Gly Glu 770 775 780		
Ile Lys Trp Leu Arg Ile Ser Ser Ser Val Lys Lys Tyr Tyr Ile His 785 790 795 800		
Asp His Phe Ile Pro Ile Glu Lys Tyr Gln Phe Ser Leu Tyr Pro Ile 805 810 815		
Phe Met Glu Gly Val Gly Lys Pro Lys Ile Ile Asn Ser Phe Thr Gln 820 825 830		
Asp Asp Ile Glu Lys His Gln Ser Asp Ala Gly Leu Tyr Val Ile Val 835 840 845		
Pro Val Ile Ile Ser Ser Ser Ile Leu Leu Leu Gly Thr Leu Leu Ile 850 855 860		
Ser His Gln Arg Met Lys Lys Leu Phe Trp Glu Asp Val Pro Asn Pro 865 870 875 880		
Lys Asn Cys Ser Trp Ala Gln Gly Pro Asn Phe Gln Lys Lys Met Pro 885 890 895		
Gly Thr Lys Glu Leu Leu Gly Gly Gly Trp Leu Thr 900 905		

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 21 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Glu Met Leu Val Lys Leu Arg Pro Thr Ser Cys Ala Pro Ala Leu Pro 1 5 10 15
Glu Val Cys Thr Met 20

International Application No: PCT/

/

MICROORGANISMS	
Optional Sheet in connection with the microorganism referred to on page <u>16</u> , lines <u>17-37</u> of the description *	
A. IDENTIFICATION OF DEPOSIT * Further deposits are identified on an additional sheet *	
Name of depositary institution * American Type Culture Collection	
Address of depositary institution (including postal code and country) * 12301 Parklawn Drive Rockville, MD 20852 US	
Date of deposit * <u>September 14, 1994</u> Accession Number * <u>75885</u>	
B. ADDITIONAL INDICATIONS * (Leave blank if not applicable). This information is continued on a separate attached sheet	
C. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE * (If no indication on any of designated States)	
D. SEPARATE FURNISHING OF INDICATIONS * (Leave blank if not applicable)	
The indications listed below will be submitted to the International Bureau later * (Specify the general nature of the indications e.g., "Accession Number of Deposit")	
E. <input type="checkbox"/> This sheet was received with the International application when filed (to be checked by the receiving Office)	
<div style="text-align: right;">_____ (Authorized Officer)</div>	
<input type="checkbox"/> The date of receipt (from the applicant) by the International Bureau *	
<div style="text-align: right;">was _____ (Authorized Officer)</div>	

Form PCT/RO/134 (January 1981)

WO 97/26523

PCT/US97/00881

International Application No: PCT/ /

Form PCT/RO/134 (cont.)

American Type Culture Collection

12301 Parklawn Drive
Rockville, MD 20852
US

Accession No.

75886

75888

75889

75974

Date of Deposit

September 14, 1994

September 14, 1994

September 14, 1994

December 14, 1994

WHAT IS CLAIMED IS:

1. A method for detecting a defective OB-R in cells comprising:
 - (a) extracting RNA from a cell population;
 - 5 (b) contacting the RNA with an oligonucleotide derived from a portion of the sequence depicted in Figure 1A-1E; and
 - (c) detecting hybridization of the RNA with the oligonucleotide.
- 10 2. The method of Claim 1 in which the cell population is obtained from the brain.
3. The method of Claim 1 in which the cell population
15 is obtained from the lung.
4. The method of Claim 1 in which the cell population is obtained from the kidney.
- 20 5. The method of Claim 1 in which the oligonucleotide is derived from nucleotide residue #2770 and beyond in the sequence depicted in Figure 1A-1E.
6. A method for treating obesity, comprising
25 administering to an individual an effective amount of an agent capable of inhibiting expression of an OB-R variant gene.
7. The method of Claim 6 in which the OB-R variant
30 gene further comprises the sequence of Figure 1A-1E or which is capable of selectively hybridizing to it.
8. The method of Claim 7 in which the agent is an antisense molecule complementary to mRNA encoded by the
35 sequence of Figure 1A-1E.

9. The method of Claim 7 in which the agent is a ribozyme molecule specific for mRNA encoded by the sequence of Figure 1A-1E.

5 10. The method of Claim 7 in which the agent is a triple helix component.

11. A method for identifying a compound capable of supplementing biological activity of leptin, comprising:

- 10 (a) incubating host cell expressing an OB-R variant with leptin;
- (b) incubating a portion of the leptin-treated cells with a test compound; and
- 15 (c) comparing activation signal in the cells treated in step (b) with cells treated in step (a);

thereby determining whether the compound augments activation of the OB-R variant by leptin.

20 12. The method of Claim 11 in which the OB-R variant is encoded by the sequence depicted in Figure 1A-1E.

25

30

35

CGC	CGC	CGC	ACG	CAG	GTG	CCC	GAG	CCC	CGG	CCC	GCG	CCC	ATC	TCT	GCC	TTC	GGT				
A	R	A	T	Q	V	P	E	P	R	P	A	P	I	S	A	F	G				
CGA	GTT	63	CCC	CCG	GAT	72	CAA	GGT	GTA	CTT	CTC	TGA	AGT	AAG	ATG	ATT	TGT	108	CAA		
R	V	G	P	P	D	Q	G	V	L	L	*	S	K	M	I	C	Q				
AAA	TTC	117	TGT	GTG	GTT	126	TTA	CAT	TGG	GAA	TTT	ATT	TAT	GTG	ATA	ACT	GCG	162	TTT		
K	F	C	V	V	L	L	H	W	E	P	I	Y	V	I	T	A	P				
AAC	TTG	171	TCA	TAT	CCA	180	ATT	ACT	CCT	TGG	AGA	TTT	AAG	TTG	TCT	TGC	ATG	CCA	216	CCA	
N	L	S	Y	P	I	T	P	W	R	F	K	L	S	C	M	P	P				
AAT	TCA	225	ACC	TAT	GAC	234	TAC	TTC	CTT	TTG	CCT	GCT	GGA	CTC	TCA	AAG	AAT	ACT	270	TCA	
N	S	T	Y	D	Y	F	L	L	P	A	G	L	S	K	N	T	S				
AAT	TGG	279	AAT	GGA	CAT	288	TAT	GAG	ACA	GCT	GTT	GAA	CCT	AAG	TTT	AAT	TCA	AGT	324	GGT	
N	S	N	G	H	Y	E	T	A	V	E	P	K	F	N	S	S	G				
ACT	CAC	333	TTT	TCT	AAC	342	TTA	TCC	AAA	GCA	ACT	TTC	CAC	TGT	TGC	TTT	CGG	AGT	378	GAG	
T	H	F	S	N	L	S	K	A	T	F	H	C	C	F	R	S	E				
CAA	GAT	387	AGA	AAC	TGC	396	TCC	TTA	TGT	GCA	GAC	AAC	ATT	GAA	GGA	AGG	ACA	TTT	432	GTT	
Q	D	R	N	C	S	L	C	A	D	N	I	E	G	R	T	F	V				
TCA	ACA	441	GTA	AAT	TCT	450	TTA	GTT	TTT	CAA	CAA	ATA	GAT	GCA	AAC	TGG	AAC	ATA	486	CAG	
S	T	V	N	S	L	V	F	Q	Q	I	D	A	N	W	N	I	Q				
TGC	TGG	495	CTA	AAA	GGA	504	GAC	TTA	AAA	TTA	TTC	ATC	TGT	TAT	GTG	GAG	TCA	TTA	540	TTT	
C	W	L	K	G	D	L	K	L	F	I	C	Y	V	E	S	L	F				
AAG	AAT	549	CTA	TTC	AGG	558	AAT	TAT	AAC	567	TAT	AAG	GTC	576	CAT	CTT	TTA	585	TAT	594	CCT
K	N	L	F	R	N	Y	N	Y	K	V	H	L	L	Y	V	L	P				
GAA	GTG	603	TTA	GAA	GAT	612	TCA	CCT	CTG	621	GTT	CCC	CAA	630	AAA	GGC	AGT	639	TTT	648	GTT
E	V	L	E	D	S	P	L	V	P	Q	K	G	S	F	Q	H	V				

Figure 1A

657	666	675	684	693	702
CAC TGC AAT TGC AGT GTT CAT GAA TGT TGT GAA TGT CTT GTG CCT GTG CCA ACA					
H C N C S V H E C C E C L V P V P T					
711	720	729	738	747	756
GCC AAA CTC AAC GAC ACT CTC CTT ATG TGT TTG AAA ATC ACA TCT GGT GGA GTA					
A K L N D T L L M C L K I T S G G V					
765	774	783	792	801	810
ATT TTC CGG TCA CCT CTA ATG TCA GTT CAG CCC ATA AAT ATG GTG AAG CCT GAT					
I F R S P L M S V Q P I N M V K P D					
819	828	837	846	855	864
CCA CCA TTA GGT TTG CAT ATG GAA ATC ACA GAT GAT GGT AAT TTA AAG ATT TCT					
P P L G L H M E I T D D G N L K I S					
873	882	891	900	909	918
TGG TCC AGC CCA CCA TTG GTA CCA TTT CCA CTT CAA TAT CAA GTG AAA TAT TCA					
W S S P P L V P F P L Q Y Q V K Y S					
927	936	945	954	963	972
GAG AAT TCT ACA ACA GTT ATC AGA GAA GCT GAC AAG ATT GTC TCA GCT ACA TCC					
E N S T T V I R E A D K I V S A T S					
981	990	999	1008	1017	1026
CTG CTA GTA GAC AGT ATA CTT CCT GGG TCT TCG TAT GAG GTT CAG GTG AGG GGC					
L L V D S I L P G S S Y E V Q V R G					
1035	1044	1053	1062	1071	1080
AAG AGA CTG GAT GGC CCA GGA ATC TGG AGT GAC TGG AGT ACT CCT COT GTC TTT					
K R L D G P G I W S D W S T P R V F					
1089	1098	1107	1116	1125	1134
ACC ACA CAA GAT GTC ATA TAC TTT CCA CCT AAA ATT CTG ACA AGT GTT GGG TCT					
T T Q D V I Y F P P K I L T S V G S					
1143	1152	1161	1170	1179	1188
AAT GTT TCT TTT CAC TCC ATC TAT AAG AAG GAA AAC AAG ATT GTT CCC TCA AAA					
N V S F H C I Y K K E N K I V P S K					
1197	1206	1215	1224	1233	1242
GAG ATT GTT TGG TGG ATG AAT TTA GCT GAG AAA ATT CCT CAA AGC CAG TAT GAT					
E I V W W M N L A E K I P Q S Q Y D					
1251	1260	1269	1278	1287	1296
GTT GTG AGT GAT CAT GTT AGC AAA GTT ACT TTT TTC AAT CTG AAT GAA ACC AAA					
V V S D H V S K V T F F N L N E T K					
1305	1314	1323	1332	1341	1350
CCT CGA GGA AAG TTT ACC TAT GAT GCA GTG TAC TGC TGC AAT GAA CAT GAA TGC					
P R G K F T Y D A V Y C C N E H E C					

Figure 1B

1359	1368	1377	1386	1395	1404
CAT CAT CGC TAT GCT GAA TTA TAT GTG ATT GAT GTC AAT ATC AAT ATC TCA TGT					
H H R Y A E L Y V I D V N I N I S C					
1413	1422	1431	1440	1449	1458
GAA ACT GAT GGG TAC TTA ACT AAA ATG ACT TGC AGA TGG TCA ACC AGT ACA ATC					
E T D G Y L T K M T C R W S T S T I					
1467	1476	1485	1494	1503	1512
CAG TCA CTT GCG GAA AGC ACT TTG CAA TTG AGG TAT CAT AGG AGC AGC CTT TAC					
Q S L A E S T L Q L R Y H R S S L Y					
1521	1530	1539	1548	1557	1566
TGT TCT GAT ATT CCA TCT ATT CAT CCC ATA TCT GAG CCC AAA GAT TGC TAT TTG					
C S D I P S I H P I S E P K D C Y L					
1575	1584	1593	1602	1611	1620
CAG AGT GAT GGT TTT TAT GAA TGC ATT TTC CAG CCA ATC TTC CTA TTA TCT GGC					
Q S D G F Y E C I F Q P I F L L S G					
1629	1638	1647	1656	1665	1674
TAC ACA ATG TGG ATT AGG ATC AAT CAC TCT CTA GGT TCA CTT GAC TCT CCA CCA					
Y T M W I R I N H S L G S L D S P P					
1683	1692	1701	1710	1719	1728
ACA TGT GTC CTT CCT GAT TCT GTG GTG AAG CCA CTG CCT CCA TCC AGT GTG AAA					
T C V L P D S V V K P L P P S S V K					
1737	1746	1755	1764	1773	1782
GCA GAA ATT ACT ATA AAC ATT GGA TTA TTG AAA ATA TCT TGG GAA AAG CCA GTC					
A E I T I N I G L L K I S W E K P V					
1791	1800	1809	1818	1827	1836
TTT CCA GAG AAT AAC CTT CAA TTC CAG ATT CGC TAT GGT TTA AGT GGA AAA GAA					
F P E N N L Q P Q I R Y G L S G K E					
1845	1854	1863	1872	1881	1890
GTA CAA TGG AAG ATG TAT GAG GTT TAT GAT GCA AAA TCA AAA TCT GTC AGT CTC					
V Q W K M Y E V Y D A K S K S V S L					
1899	1908	1917	1926	1935	1944
CCA GTT CCA GAC TTG TGT GCA GTC TAT GCT GTT CAG GTG CGC TGT AAG AGG CTA					
P V P D L C A V Y A V Q V R C K R L					
1953	1962	1971	1980	1989	1998
GAT GGA CTG GGA TAT TGG AGT AAT TGG AGC AAT CCA GCC TAC ACA GTT GTC ATG					
D G L G Y W S N W S N P A Y T V V M					
2007	2016	2025	2034	2043	2052
GAT ATA AAA GTT CCT ATG AGA GGA CCT GAA TTT TGG AGA ATA ATT AAT GGA GAT					
D I K V P M R G P E F W R I I N G D					

Figure 1C

2061	2070	2079	2088	2097	2106
ACT ATG AAA AAG GAG AAA AAT GTC ACT TTA CTT TGG AAG CCC CTG ATG AAA AAT					
T M K K E K N V T L L W K P L M K N					
2115	2124	2133	2142	2151	2160
GAC TCA TTG TGC AGT GTT CAG AGA TAT GTG ATA AAC CAT CAT ACT TCC TGC AAT					
D S L C S V Q R Y V I N H H T S C N					
2169	2178	2187	2196	2205	2214
GGA ACA TGG TCA GAA GAT GTG GGA AAT CAC ACG AAA TTC ACT TTC CTG TGG ACA					
G T W S E D V G N H T K F T F L W T					
2223	2232	2241	2250	2259	2268
GAG CAA GCA CAT ACT GTT ACG GTT CTG GCC ATC AAT TCA ATT GGT GCT TCT GTT					
E Q A H T V T V L A I N S I G A S V					
2277	2286	2295	2304	2313	2322
GCA AAT TTT AAT TTA ACC TTT TCA TGG OCT ATG AGC AAA GTA AAT ATC GTG CAG					
A N F N L T F S W P M S K V N I V Q					
2331	2340	2349	2358	2367	2376
TCA CTC AGT GCT TAT OCT TTA AAC AGC AGT TGT GTG ATT GTT TCC TGG ATA CTA					
S L S A Y P L N S S C V I V S W I L					
2385	2394	2403	2412	2421	2430
TCA CCC AGT GAT TAC AAG CTA ATG TAT TTT ATT ATT GAG TGG AAA AAT CTT AAT					
S P S D Y K L M Y F I I E W K N L N					
2439	2448	2457	2466	2475	2484
GAA GAT GGT GAA ATA AAA TGG CTT AGA ATC TCT TCA TCT GTT AAG AAG TAT TAT					
E D G E I K W L R I S S S V K K Y Y					
2493	2502	2511	2520	2529	2538
ATC CAT GAT CAT TTT ATC CCC ATT GAG AAG TAC CAG TTC AGT CTT TAC CCA ATA					
I H D H F I P I E K Y Q F S L Y P I					
2547	2556	2565	2574	2583	2592
TTT ATG GAA GGA GTG GGA AAA CCA AAG ATA ATT AAT AGT TTC ACT CAA GAT GAT					
F M E G V G K P K I I N S F T Q D D					
2601	2610	2619	2628	2637	2646
ATT GAA AAA CAC CAG AGT GAT GCA GGT TTA TAT GTA ATT GTG CCA GTA ATT ATT					
I E K H Q S D A G L Y V I V P V I I					
2655	2664	2673	2682	2691	2700
TCC TCT TCC ATC TTA TTG CTT GGA ACA TTA TTA ATA TCA CAC CAA AGA ATG AAA					
S S S I L L L G T L L I S H Q R M K					
2709	2718	2727	2736	2745	2754
AAG CTA TTT TGG GAA GAT GTT CCG AAC CCC AAG AAT TGT TCC TGG CCA CAA GGA					
K L F W E D V P N P K N C S W A Q G					

2763	2772	2781	2790	2799	2808
CTT AAT TTT	CAG AAG AAA	ATG OCT GGC	ACA AAG GAA	CTA CTG GGT	GGA GGT TGG
---	---	---	---	---	---
L N F	Q K K	M P G	T K E	L L G	G G W
2817	2826	2835	2844	2853	2862
TTG ACT TAG	GAA ATG CTT	GTG AAG CTA	OCT CCT ACC	TCG TGC GCA	CCT GCT CTC
---	---	---	---	---	---
L T *	E M L	V K L	R P T	S C A	P A L
2871	2880				
CCT GAG GTG	TGC ACA ATG	3'			
---	---				
P E V	C T M				